Evaluating conservation units using network analysis: a sea duck case study

Juliet S Lamb^{1,2*†}, Clara Cooper-Mullin¹, Scott G Gilliland³, Alicia M Berlin⁴, Timothy D Bowman⁵, W Sean Boyd⁶, Susan EW De La Cruz⁷, Daniel Esler⁸, Joseph R Evenson⁹, Paul Flint⁸, Christine Lepage¹⁰, Dustin E Meattey¹¹, Jason E Osenkowski², Peter WC Paton¹, Matthew C Perry⁴, Dan Rosenberg¹², Jean-Pierre L Savard¹³, Lucas Savoy¹¹, Jason Schamber¹², David H Ward⁸, John Y Takekawa¹⁴, and Scott R McWilliams¹

Conserving migratory wildlife requires understanding how groups of individuals interact across seasons and landscapes. Telemetry reveals individual movements at large spatiotemporal scales; however, using movement data to define conservation units requires scaling up from individual movements to species- and community-level patterns. We developed a framework to define flyways and identify important sites from telemetry data and applied it to long-term, range-wide tracking data from three species (640 individuals) of sea ducks: namely, North American scoters (*Melanitta* spp). Our network of 88 nodes included both multispecies hotspots and areas uniquely important to individual species. We found limited spatial overlap between scoters wintering on the Atlantic and Pacific coasts of North America, with differing connectivity patterns between coasts. Finally, we identified four multispecies conservation units that did not correspond to traditional management flyways. From this approach, we show how individual movements can be used to quantify range-wide connectivity of migratory species and reveal gaps in conservation strategies.

Front Ecol Environ 2024; 0(0): e2648, doi:<u>10.1002/fee.2648</u>

Migratory animals can cover vast distances, with annualcycle movements spanning biogeographic and geopolitical boundaries. Migratory movements expose species to varying ecological conditions, stressors, and management regimes, making such species challenging to conserve (Dunn *et al.* 2019). In recent decades, management of migratory wildlife has focused on coordinated regional, international, and intercontinental planning efforts (Boere and Stroud 2006). Nevertheless, many species of migratory birds have declined sharply in abundance over the past 50 years despite conservation efforts, even among taxa that have experienced net population increases (Rosenberg *et al.* 2019). This outcome suggests that current conservation measures are inadequate for addressing factors driving trends in migratory populations.

¹Department of Natural Resources Science, University of Rhode Island, *Kingston, RI;*[†]*current address: The Nature Conservancy, Cold Spring* Harbor, NY^{*}(juliet.lamb@tnc.org); ²Rhode Island Department of Environmental Management, West Kingston, RI; ³Canadian Wildlife Service, Sackville, Canada; ⁴US Geological Survey, Eastern Ecological Research Center, Laurel, MD; ⁵Sea Duck Joint Venture, US Fish and Wildlife Service, Anchorage, AK; 6Science & Technology Branch, Environment and Climate Change Canada, Delta, Canada; ⁷US Geological Survey, Western Ecological Research Center, Moffett Field, CA; ⁸US Geological Survey, Alaska Science Center, Anchorage, AK; ⁹Washington Department of Fish and Wildlife, Olympia, WA; ¹⁰Canadian Wildlife Service, Quebec City, Canada; ¹¹Biodiversity Research Institute, Portland, ME; ¹²Alaska Department of Fish and Game, Anchorage, AK; ¹³Science & Technology Branch, Environment and Climate Change Canada, Quebec City, Canada; ¹⁴Suisun Resource Conservation District, Suisun City, CA

The challenge of protecting migratory species is compounded by the complexity of defining manageable populations and identifying critical habitat areas across species ranges. Indeed, interlinked single-species populations and multispecies communities may overlap during parts of the annual cycle but diverge widely in others (Webster *et al.* 2002). In these cases, actions targeting specific locations or time periods may be insufficient if limiting factors occur in other locations or periods (Newton 2004). Likewise, interventions that benefit stable segments of the population may not reverse declines in other subpopulations, even if the groups are sympatric during parts of the year (Kramer *et al.* 2018). Identifying and targeting causes of decline therefore requires first quantifying connectivity among population units over time and space (Marra *et al.* 2015).

Advances in tracking technologies have transformed the science of animal movement from single-site studies with small sample sizes to large, continental- and global-scale repositories. These changes make it possible to follow numerous individuals across the annual cycle to understand how breeding, non-breeding, and migratory habitats are interlinked (eg Meattey *et al.* 2018; Lamb *et al.* 2020). However, analysis of telemetry data often emphasizes habitat use rather than population- and community-level connectivity patterns. As larger datasets for multiple species become available, methods that combine telemetry data across individuals, sites, and species are required to identify annual-cycle linkages and optimize conservation actions.

One promising option for integrating large telemetry datasets is network analysis (Xu *et al.* 2020): a branch of graph theory that quantitatively assesses pairwise relationships among actors, objects, or locations, and uses these relationships to describe the properties of the overall system and its components. Networks have broad applications in biology and ecology, including in genetics (Nacu *et al.* 2007), social-systems ecology (Hasenjager and Dugatkin 2015), cognitive processes (Gray *et al.* 2014), and machine learning (Lek and Guégan 1999). To date, network analysis of animal movement has typically been used to examine relationships among discrete locations (eg Iwamura *et al.* 2013). However, it can also be a powerful tool for using individual movement data to identify key sites and their interconnections in landscape-level migratory networks without predefined habitat patches (Lamb *et al.* 2019).

Our goal was to use network modeling to document community-level, continental-scale migratory connectivity from satellite telemetry data. We evaluated our method based on tracking data for three congeneric species of North American sea ducks-black scoter (Melanitta americana), surf scoter (Melanitta perspicillata), and white-winged scoter (Melanitta deglandi)—across their North American ranges. In contrast to other harvested waterfowl, sea ducks often have uncertain population trajectories, and North American scoters breed in remote boreal habitats where their reproductive success is difficult to measure (Bowman et al. 2015; Rosenberg et al. 2019). In contrast, their multistage migrations include coastal habitats with intense human activity. Sea ducks are managed using a flyway system developed for mid-continental waterfowl (Nichols et al. 1995), which was not designed to capture spatial population structure for sea ducks. Our primary objectives were to identify locations associated with singleand multispecies annual-cycle connectivity at the continental scale and assess alignment of management units with spatial structure of scoter populations.

Methods

From October to May for 21 consecutive years (1998–2019; target species and locations varied among years), we captured scoters using decoys and net guns, night-lighting, floating mist nets, or gill nets at multiple sites in eastern and western North America (Appendix S1: Table S1). We implanted 26–50-g platform terminal transmitters (PTTs; Microwave Telemetry Inc, Columbia, Maryland, or Telonics, Mesa, Arizona) in 782 individual scoters, of which 538 (143–235 transmitters per species) collected multi-season data suitable for our analysis (Appendix S1: Table S1). Sample sizes for all species were slightly higher in the eastern region (58% of all retained transmitters; 54–60% per species). Capture and implantation methods are fully described in De La Cruz *et al.* (2009), Lamb *et al.* (2019), and Bowman *et al.* (2021).

Transmitters collected location data approximately once every 3–6 days (see De La Cruz *et al.* [2009] and Lamb *et al.* [2019] for duty cycles). Locations were derived from the Doppler shift in transmitter signals, received by polar-orbiting satellites, and compiled through the CLS (Collection & Location by Satellite) America processing center in Lanham, Maryland. We regularly downloaded transmission data, which were filtered through a hybrid Douglas Argos Filter (Douglas *et al.* 2012) to retain the single location with the highest accuracy from each duty cycle and reduce redundant positional information.

We estimated regular movement trajectories using a switching state-space model (Jonsen *et al.* 2005) to account for location error and temporal gaps. Tracks were interpolated to one location per day and daily locations were classified based on their probability of assignment to either resident (shorter step distances, frequent directional change) or transient (longer distances, consistent direction) behavioral states (see Lamb *et al.* [2019] for full model parameterization and state characteristics). We assumed resident birds were actively using habitat, whereas transient birds were flying through. We defined a residency period as ≥ 2 interpolated resident locations separated before and after by ≥ 2 transient locations. All models were run using the *bsam* package (Jonsen *et al.* 2005) in R (R Core Team 2018).

Within each residency period, we calculated the occupancy duration and geographic centroid of all locations. Each centroid was assigned to a season based on its phenology within the annual cycle following De La Cruz *et al.* (2009): wintering (November–February), spring migration (February–May), breeding (May–July; inland), or molt and fall migration (May– November for non-breeders and males, July–November for breeding females; primarily coastal). We omitted each individual's first habitat centroid to avoid inflating the relative importance of capture locations and exclude abnormal post-capture movements (Lamb *et al.* 2020).

To define groups of habitat centroids, we selected the optimal number of clusters using the R package *NbClust* (Charrad *et al.* 2014), varying the number of clusters from 20 to 200 and choosing the consensus solution across multiple algorithms. Using the resulting clusters (nodes) and between-cluster movements (edges), we constructed single- and multispecies networks using the R package *igraph* (Csárdi *et al.* 2015). Network analysis evaluates pairwise relationships (here, individual movements) among entities (here, locations) to assess the structure of the community and the roles of its components. We inversely weighted each habitat centroid or edge by the proportion of centroids or edges from that species and region to ensure that each species and region contributed equally to the final network.

Following network construction, we evaluated four nodespecific importance metrics using the *igraph* package and Conefor 2.6 software (Saura and Torné 2009): outdegree centrality (number of outgoing edges), indegree centrality (number of incoming edges), betweenness centrality (number of pathways including the target node), and connectivity (dPC [change in probability of connectivity]; here, the change in connectivity after removing the target node) (Appendix S1: Figure S1). To compare interspecific network use and identify opportunities for shared threats or benefits among scoter species, we calculated centrality values across the multispecies network and for each species individually. We tested the sensitivity of our results to weighting factors by comparing several alternative weighting schemes, including by individual, capture site, duration of use, and combinations of factors (Appendix S1: Figure S2). We found that site rankings were similar regardless of weighting factors (coefficient of variation [CV] of rank: mean = 0.27; range = 0.00-1.64).

To identify groups of linked locations, we assessed network modularity, or the extent to which subgroups of nodes (modules) in a network are more closely connected with one another than expected if linkages were random. We identified modules using propagating labels in *igraph* and evaluated results using the Girvan-Newman algorithm (Appendix S1: Figure S1; Girvan and Newman 2002), which does not require a priori knowledge of the expected number of modules. Under this system, a network with weak migratory connectivity (ie a network in which individuals from different breeding areas are evenly mixed at wintering sites; Webster et al. 2002) would score closer to zero, whereas strong connectivity (ie individuals from different breeding sites also segregate on wintering grounds) would score closer to one. A score of one would require all nodes within a module to directly interconnect, which we did not expect because sea ducks use stopover nodes between wintering and breeding. We therefore considered modularity scores closer to fully interconnected modules than random linkages (ie \ge 0.5) to represent strong connectivity.

Results

The multispecies North American scoter network consisted of 88 nodes (Appendix S1: Figure S3 and Table S2). Of these, 44 (50%) were used only by scoters captured in eastern North America, 29 (33%) only by scoters captured in western North America, and 15 (17%) by scoters from both groups during spring migration, fall migration, and breeding (Figure 1). Scoters in eastern North America used distinct nodes for wintering, spring migration, breeding, and fall migration/molt, whereas scoters from western North America utilized nodes during multiple seasons. The northernmost terrestrial breeding nodes also included marine coastlines used during molt (Figure 1; Appendix S1: Figure S3).

Several coastal nodes used during migration had high values across all centrality metrics (Figure 2; Appendix S1: Table S2), indicating that they were widely used by tracked individuals and served as points of connection across the



Figure 1. (a) Multispecies annual-cycle network for North American black scoter (*Melanitta americana*), surf scoter (*Melanitta perspicillata*), and white-winged scoter (*Melanitta deglandi*) showing seasonal use (blue = winter, green = spring migration, red = breeding, orange = fall migration), 1998–2019. Node sizes correspond to relative intensity of use, with the node size key (nested circles) depicting numbers of centroids for small-, medium-, and large-diameter nodes. Inset maps depict areas of overlap between individuals from western (dark purple) and eastern (dark pink) capture sites for (b) white-winged scoter and (c) surf scoter. Eastern and western black scoter populations did not overlap.

migratory network. Indegree and outdegree centrality and connectivity were highest at coastal staging nodes and wintering nodes on both coasts (Figure 2, a and b, d), and betweenness centrality was also high at breeding and molting nodes where eastern and western scoters overlapped (Figure 2c). Species-specific centrality was comparatively higher in coastal spring and fall migration nodes for black scoters, and in coastal wintering and inland breeding nodes for surf scoters and white-winged scoters (Appendix S1: Figure S4).

The multispecies network was highly modular and contained four interlinked modules (one in the east and three in the west), which did not correspond to existing management units (Figure 3a). Modularity scores were high for singlespecies networks (Figure 3, b–d), indicating distinct subunits within the continental population for all scoter species. The



Figure 2. Multispecies values for (a) indegree centrality, (b) outdegree centrality, (c) betweenness centrality, and (d) connectivity for North American scoters, 1998–2019. For complete descriptions of the four network centrality metrics, see Appendix S1: Figure S1.

black scoter network consisted of isolated eastern and western modules (Figure 3c), whereas the surf scoter network contained two interlinked modules (Figure 3b) and the whitewinged scoter network contained three modules (Figure 3d). Modularity scores were similarly high in all three species (~0.5), suggesting stronger linkages within than between modules. Boundaries between eastern and western subgroups were around 106°W for surf scoters, and 113°W for black scoters and white-winged scoters.

Discussion

Integrating range-wide telemetry data into a spatial network allowed us to assess the relative importance of sites to annualcycle movement of migratory birds and identify high-value locations. By measuring and describing patterns of migratory connectivity across the ranges of three scoter species, we highlighted key similarities and differences in network structure among species and regions. In addition, scoter networks were highly modular and spanned multiple management units, highlighting the need for regional coordination.

Our study adds to a growing body of applications of network analysis to movements of migratory species and communities (eg Iwamura *et al.* 2013; Xu *et al.* 2020; Donnelly *et al.* 2021). Like these studies, our application of network analysis reveals the high importance of migratory stopover

Front Ecol Environ doi:10.1002/fee.2648

sites to overall network connectivity and allows explicit prioritization of sites based on their relationships to other locations within migratory networks. While previous studies often focused on predefined networks of habitat patches, we extended these analyses by using telemetry data rather than habitat to define nodes. Our approach can be flexibly applied to species for which key habitats are not defined a priori. However, it is sensitive to decisions such as clustering methods and weighting, which must be carefully selected. In addition, generating a representative network requires distributing transmitters throughout the population(s) of interest. This study benefited from a coordinated effort to deploy transmitters throughout the populations of interest, resulting in comprehensive coverage of scoter species' ranges (Pearce et al. 2019) with sample sizes sufficient to characterize seasonal distributions (ie at least 80-130 individuals per species; Roberts et al. 2018). However, low-density wintering areas (eg the Gulf of Mexico for black scoters) were unsampled and are underrepresented in the network. Less common migratory routes could also be underrepresented by chance in our network, which could result in underestimation of connectivity along these routes. Staging sites exhibited high in- and outdegree centrality and population mixing, making them useful targets for increasing sample representativeness, but could still exclude alternative migratory routes. If sampling design is carefully considered, however, our approach offers a straightforward,

repeatable, and transparent mechanism for translating telemetry data into a landscape framework.

Our analysis identified important sites for multispecies connectivity, which could be valuable targets for maximizing conservation benefits across species. For scoters, coastal staging sites in Nantucket Sound, the St Lawrence River, the Salish Sea, and southeast and southwest Alaska had high in- and outdegree centrality, suggesting intermixing of individuals from various breeding and wintering sites. In addition, the white-winged scoter and surf scoter networks each included different breeding nodes with high betweenness centrality values used by individuals from eastern and western wintering areas. Despite similarities in their ranges, migratory patterns, and habitat requirements, however, the three North American scoter species showed differences in breeding and nonbreeding habitat selection (Takekawa et al. 2011; Lamb et al. 2020). Correspondingly, we observed among-species differences in importance of shared sites. In southern Alaska, for instance, Prince William Sound was relatively more central to the whitewinged scoter network, whereas southeast Alaska was more central to the surf scoter network. Thus, while it may be optimal to conserve sites with high importance values across multiple species, maintaining a strong migratory network also requires including sites with unique single-species importance.

Our results also highlight regional and species-specific differences in migratory connectivity patterns. While migration of eastern scoters was concentrated along relatively uniform routes consisting of northsouth movements to stopover and staging sites followed by east-west breeding or molting migrations, migration of western scoters was diffuse over many routes, with direct movements between inland staging and breeding sites and coastal molt and wintering areas. Eastern and western white-winged scoters overlapped over a broad longitudinal range, from Hudson Bay to the Northwest

Territories; surf scoters overlapped only in a few nodes; and black scoters did not spatially overlap at all. Furthermore, western black scoters migrated shorter distances than eastern black scoters and the other two scoter species, with a unique migration from multispecies wintering areas in southeast Alaska, British Columbia, and the Salish Sea to southwestern Alaska. Together, these differences suggest that effectiveness of



Figure 3. Network modularity for North American scoters tracked between 1998 and 2019 for (a) all three scoter species combined, (b) surf scoters only, (c) black scoters only, and (d) whitewinged scoters only. Colors indicate groups of nodes (ie modules) that are more interconnected with one another than with the overall network. Modularity values (Girvan-Newman scores) ranged from 0 (random) to 1 (complete interconnection within modules and no connections between modules); we considered values >0.5 to represent high modularity. Dashed lines in (a) show boundaries of traditional waterfowl flyways (Pacific, Central, Mississippi, and Atlantic) in the US and Canada.

multispecies conservation planning may vary by region, with western scoters requiring a more species-specific approach.

Individual movements delineated four groups of scoters that traverse and subdivide existing flyways in North America. All species occupied a single eastern module spanning the Atlantic, Mississippi, and Central flyways (supporting Lamb *et al.* 2019). Three multispecies western modules divided along

a northwest-southeast axis, each spanning the Pacific and Central flyways. Number and configuration of modules varied by species in the western portion of their ranges, with less multispecies overlap. Our results suggest that coordination across North American flyways—and, occasionally, subdivision within flyways—may be needed to distribute management actions across annual-cycle habitats for scoters or species with similarly complex migratory patterns (eg sandhill crane [*Antigone canadensis*]; Donnelly *et al.* 2021). Using individual movements to define management units could offer opportunities to link population trends to relevant demographic rates and external drivers across the annual cycle.

Although our application of network analysis focused on a specific group of sea duck species, similar approaches could be applied to other species and communities of migratory wildlife for which large-scale tracking datasets are available. For example, multispecies tracking data have been compiled across a suite of Arctic wildlife (Davidson *et al.* 2020) and could be analyzed in a network structure to compare modularity among species and to identify migratory pathways and locations vulnerable to habitat loss under future climate scenarios. Telemetry data could also be used to construct or validate spatial networks for identifying populations of mobile wildlife affected by barriers along migratory corridors, such as turtles impacted by road crossings (Pereira *et al.* 2011) or bats exposed to wind energy installations (Erickson *et al.* 2016).

Conclusions

For North American scoters, our analysis revealed interconnected subgroups that transcend geopolitical management boundaries. Sea ducks spend most of the annual cycle in coastal non-breeding habitats along the Atlantic and Pacific coasts, providing opportunities for conservation and harvest management within these units; however, wintering areas within the same management flyway may be connected to different migratory and breeding habitats. Connectivity-based analyses offer mechanisms for contextualizing varying population trajectories within existing management units and optimizing management decisions across the annual cycle. In addition, although eastern and western populations of surf scoters and white-winged scoters are not genetically distinct (Sonsthagen et al. 2019), we found limited spatial overlap between these populations, which could result in separate demographic trajectories. Thus, management and monitoring of eastern and western scoters as distinct subpopulations may be warranted despite genetic overlap.

For scoters and other migratory wildlife, movement networks like ours provide a valuable starting point for further analysis. Integrating survey and demographic data into networks (Studds *et al.* 2017) can help identify differential population trends among modules and assess how vital rates associated with stage-specific habitats propagate throughout the network. Furthermore, assessing relationships between habitat characteristics and node centrality (Donnelly *et al.* 2021) and incorporating predictive modeling provide a basis for evaluating how future changes in habitat will impact migratory movements, and which nodes and pathways may become more important to migratory connectivity under climate warming and habitat loss scenarios. These and other extensions of network analyses could provide valuable insights for conservation of long-distance migrants in a changing world.

Acknowledgements

This work was permitted by institutional Animal Care and Use Committees; the Canadian Wildlife Service, US Fish and Wildlife Service, and US Bird Banding Laboratory; and multiple states and territories. Funding and in-kind support was provided by Sea Duck Joint Venture, Environment and Climate Change Canada, Bureau of Ocean Energy Management, US Department of Energy, US Geological Survey, Natural Sciences and Engineering Research Council of Canada, Alaska Department of Fish and Game, Kodiak National Wildlife Refuge, Exxon Valdez Oil Spill Trustee Council, Alaska SeaLife Center, US Coast Guard, Bird Studies Canada, Ducks Unlimited Inc, Wildlife Restoration, University of Rhode Island, Rhode Island Department of Environmental Management, CALFED Ecosystem Restoration Program, University of California, Simon Fraser University, and National Aeronautics and Space Administration Signals of Spring. Veterinarians were C Brown, S Ford, S Gibbs, S Lair, S Larrat, RLeNet, CLeRochais, J Leach, M McAdie, D Mulcahy, G Olsen, A Santamaria-Bouvier, C Scala, and P Tuomi. A Gilbert and D Douglas compiled datasets and recovered information, and many collaborators (listed in De La Cruz et al. [2009]; Lamb et al. [2019]; Lepage et al. [2020]; Bowman et al. [2021]) provided field, lab, and design support. Any use of trade, firm, or product names is for descriptive purposes only and does not imply endorsement by the US Government.

Data Availability Statement

Data and novel code used in this manuscript are available on Zenodo (https://doi.org/10.5281/zenodo.10137095).

References

- Boere GC and Stroud DA. 2006. The flyway concept: what it is and what it isn't. In: Boere GC, Galbraith CA, and Stroud DA (Eds). Waterbirds around the world: a global overview of the conservation, management and research of the world's waterbird flyways. Edinburgh, UK: The Stationery Office.
- Bowman TD, Gilliland SG, Schamber JL, *et al.* 2021. Strong evidence for two disjunct populations of black scoters (*Melanitta americana*) in North America. *Wildfowl* **71**: 179–92.
- Bowman TD, Silverman ED, Gilliland SG, et al. 2015. Status and trends of North American sea ducks. In: Savard J-PL, Derksen DV,

Esler D, and Eadie JM (Eds). Ecology and conservation of North American sea ducks. Boca Raton, FL: CRC Press.

- Charrad M, Ghazzali N, Boiteau V, *et al.* 2014. NbClust: an R package for determining the relevant number of clusters in a data set. *J Stat Software* **61**: 1–36.
- Csárdi G, Nepusz T, Traag V, *et al.* 2015. igraph: network analysis and visualization. Vienna, Austria: R Foundation for Statistical Computing.
- Davidson SC, Bohrer G, Gurarie E, *et al.* 2020. Ecological insights from three decades of animal movement tracking across a changing Arctic. *Science* **370**: 712–15.
- De La Cruz SE, Takekawa JY, Wilson MT, *et al.* 2009. Spring migration routes and chronology of surf scoters (*Melanitta perspicillata*): a synthesis of Pacific coast studies. *Can J Zool* **87**: 1069–86.
- Donnelly JP, King SL, Knetter J, *et al.* 2021. Migration efficiency sustains connectivity across agroecological networks supporting sandhill crane migration. *Ecosphere* **12**: e03543.
- Douglas DC, Weinzierl R, Davidson S, *et al.* 2012. Moderating Argos location errors in animal tracking data. *Methods Ecol Evol* **3**: 999–1007.
- Dunn DC, Harrison AL, Curtice C, et al. 2019. The importance of migratory connectivity for global ocean policy. P Roy Soc B-Biol Sci 286: 20191472.
- Erickson RA, Thogmartin WE, Diffendorfer JE, *et al.* 2016. Effects of wind energy generation and white-nose syndrome on the viability of the Indiana bat. *PeerJ* **4**: e2830.
- Girvan M and Newman ME. 2002. Community structure in social and biological networks. *P Natl Acad Sci USA* **99**: 7821–26.
- Gray SA, Zanre E, and Gray SR. 2014. Fuzzy cognitive maps as representations of mental models and group beliefs. In: Papageorgiou EI (Ed). Fuzzy cognitive maps for applied sciences and engineering. Heidelburg, Germany: Springer.
- Hasenjager MJ and Dugatkin LA. 2015. Social network analysis in behavioral ecology. *Adv Study Behav* 47: 39–114.
- Iwamura T, Possingham HP, Chadès I, et al. 2013. Migratory connectivity magnifies the consequences of habitat loss from sealevel rise for shorebird populations. P Roy Soc B-Biol Sci 280: 20130325.
- Jonsen ID, Flemming JM, and Myers RA. 2005. Robust state-space modeling of animal movement data. *Ecology* **86**: 2874–80.
- Kramer GR, Andersen DE, Buehler DA, *et al.* 2018. Population trends in *Vermivora* warblers are linked to strong migratory connectivity. *P Natl Acad Sci USA* **115**: E3192–200.
- Lamb JS, Paton PW, Osenkowski JE, *et al.* 2019. Spatially explicit network analysis reveals multi-species annual cycle movement patterns of sea ducks. *Ecol Appl* **29**: e01919.
- Lamb JS, Paton PW, Osenkowski JE, *et al.* 2020. Implanted satellite transmitters affect sea duck movement patterns at short and long timescales. *Condor* **122**: duaa029.
- Lek S and Guégan JF. 1999. Artificial neural networks as a tool in ecological modelling, an introduction. *Ecol Model* **120**: 65–73.

- Lepage C, Savard J-PL, and Gilliland SG. 2020. Spatial ecology of whitewinged scoters (*Melanitta deglandi*) in eastern North America: a multi-year perspective. *Waterbirds* 43: 147–62.
- Marra PP, Cohen EB, Loss SR, *et al.* 2015. A call for full annual cycle research in animal ecology. *Biol Lett-UK* **11**: 20150552.
- Meattey DE, McWilliams SR, Paton PWC, *et al.* 2018. Annual cycle of white-winged scoters (*Melanitta fusca*) in eastern North America: migratory phenology, population delineation, and connectivity. *Can J Zool* **96**: 1353–65.
- Nacu Ş, Critchley-Thorne R, Lee P, *et al.* 2007. Gene expression network analysis and applications to immunology. *Bioinformatics* 23: 850–58.
- Newton I. 2004. Population limitation in migrants. Ibis 146: 197-226.
- Nichols JD, Johnson FA, and Williams BK. 1995. Managing North American waterfowl in the face of uncertainty. *Ann Rev Ecol Syst* **26**: 177–99.
- Pearce JM, Flint PL, Whalen ME, et al. 2019. Visualizing populations of North American sea ducks – maps to guide research and management planning. Reston, VA: US Geological Survey.
- Pereira M, Segurado P, and Neves N. 2011. Using spatial network structure in landscape management and planning: a case study with pond turtles. *Landscape Urban Plan* **100**: 67–76.
- R Core Team. 2018. R: a language and environment for statistical computing. Vienna, Austria: R Foundation for Statistical Computing.
- Roberts A, Silverman E, and Gifford S. 2018. Sample size considerations for satellite telemetry and animal distributions. *J Wildlife Manage* **82**: 1536–44.
- Rosenberg KV, Dokter AM, Blancher PJ, *et al.* 2019. Decline of the North American avifauna. *Science* **366**: 120–24.
- Saura S and Torné J. 2009. Conefor Sensinode 2.2: a software package for quantifying the importance of habitat patches for landscape connectivity. *Env Model Softw* **24**: 135–39.
- Sonsthagen SA, Wilson RE, Lavretsky P, *et al.* 2019. Coast to coast: high genomic connectivity in North American scoters. *Ecol Evol* **9**: 7246–61.
- Studds CE, Kendall BE, Murray NJ, *et al.* 2017. Rapid population decline in migratory shorebirds relying on Yellow Sea tidal mudflats as stopover sites. *Nat Commun* **8**: 14895.
- Takekawa JY, De La Cruz SW, Wilson MT, *et al.* 2011. Breeding distribution and ecology of Pacific coast surf scoters. In: Wells JV (Ed). Boreal birds of North America: a hemispheric view of their conservation links and significance. Berkeley, CA: University of California Press.
- Webster MS, Marra PP, Haig SM, *et al.* 2002. Links between worlds: unraveling migratory connectivity. *Trends Ecol Evol* **17**: 76–83.
- Xu Y, Si Y, Takekawa J, *et al.* 2020. A network approach to prioritize conservation efforts for migratory birds. *Conserv Biol* **34**: 416–26.

Supporting Information

Additional material can be found online at http://onlinelibrary.wiley.com/doi/10.1002/fee.2648/suppinfo Appendix S1

Supplement to:

Evaluating conservation units using network analysis: a sea duck case study

JS Lamb et al.

Published in *Frontiers in Ecology and the Environment*

JS Lamb et al. - Supporting Information

(A) OUTDEGREE CENTRALITY

(Freeman 1979): Number of outgoing connections from a node. Higher values within our measured range indicate points of overlap preceding dispersal events; e.g., a spring staging site used by individuals that then travel to different breeding areas.



(Girvan and Newman 2002): Degree (0-1) to which the network is divided into subgroups of interconnected nodes. Values closer to one indicate a higher degree of separation among subgroups; e.g., individuals using the same breeding areas also use the same wintering sites.



Figure S1. Complete descriptions of network centrality metrics used to calculate node importance. (a) Outdegree centrality (multispecies network range: 1.75–55.44). (b) Indegree centrality (multispecies network range: 3.02–71.89). (c) Betweenness centrality (multispecies network range: 0–1710). (d) Connectivity (dPC; multispecies network range: 0.01–10.63). (e) Modularity.

Freeman LC. 1977. A set of measures of centrality based on betweenness. *Sociometry* **40**: 35–41. Freeman LC. 1979. Centrality in networks: I. conceptual clarification. *Soc Networks* **1**: 215–39. Girvan M and Newman ME. 2002. Community structure in social and biological networks. *P*

Natl Acad Sci USA 99: 7821–26.

Saura S and Pascual-Hortal L. 2007. A new habitat availability index to integrate connectivity in landscape conservation planning: comparison with existing indices and application to a case study. *Landscape Urban Plan* **83**: 91–103.





Figure S2. Standardized values for indegree centrality (top row), outdegree centrality (second row), betweenness centrality (third row), and change in probability of connectivity (bottom row) obtained using different weighting factors. Yellower and more opaque

colors represent higher centrality values. The leftmost column represents the selected weighting scheme in which each species and region contribute equally to the network. For each alternative scheme the absolute value of the mean change in scaled node weights (ie 1 = one standard deviation) under the alternative scheme compared to the species and region weighting is shown in the lower left corner of the panel.

JS Lamb et al. – Supporting Information



Figure S3. Habitat nodes (n = 88) determined from unsupervised clustering of satellite telemetry data from North American scoters, 1998–2019. Full location descriptions are provided in Table S2.

JS Lamb et al. - Supporting Information



Figure S4. Network values for betweenness centrality, connectivity (dPC), indegree centrality, and outdegree centrality for North American (a–d) black scoters (*Melanitta americana*), (e–h) surf scoters (*Melanitta perspicillata*), and (i–l) white-winged scoters (*Melanitta deglandi*), 1998–2019.

JS Lamb *et al.* – Supporting Information

Table S1. Transmitters deployed on scoters in North America from 1998 to 2019 by location, species, year, and season

		Black Scoter		Surf Scoter		White-winged Scoter				Deployment	Deployment	
Location	Region	Deployed	Retained	Deployed	Retained	Deployed	Retained	Latitude	Longitude	season	years	
NEWFOUNDLAND & LABRA	DOR											
Voiseys Bay	Eastern	0	0	14	8	0	0	56.28	-62.03	Fall molt	2006	
QUEBEC												
Forestville, Québec (St. Lawrence River)	Eastern	0	0	79	41	60	52	48.40	-69.20	Fall molt	2010 – 2016	
Chaleur Bay, New Brunswick and Québec	Eastern	93	79	6	3	0	0	47.87	-65.40	Spring migration	2002 – 2005 2009 – 2010	
GULF OF MAINE												
Merrimack River	Eastern	0	0	0	0	5	3	44.73	-66.49	Winter	2011 – 2013	
SOUTHERN NEW ENGLAND	/ NEW YO	RK										
Cape Cod Bay	Eastern	0	0	0	0	22	21	41.63	-70.21	Winter	2015 – 2016	
Rhode Island	Eastern	17	8	1	1	1	1	41.45	-71.33	Winter	2010 – 2011	
Long Island Sound	Eastern	0	0	0	0	7	5	41.13	-71.79	Winter	2014 – 2016	
GREAT LAKES												
Lake Ontario	Eastern	0	0	0	0	2	1	43.29	-79.48	Winter	2013	
MID-ATLANTIC												
Chesapeake Bay	Eastern	2	2	70	57	0	0	38.57	-76.26	Winter	2001 – 2005 2011 – 2015	
Delaware Bay	Eastern	0	0	16	11	0	0	38.80	-75.16	Winter	2013 – 2015	
SOUTH ATLANTIC BIGHT												
Pamlico Sound	Eastern	0	0	21	18	0	0	35.28	-76.33	Winter	2012 – 2015	
ALASKA												
Aropuk Lake	Western	20	15	0	0	0	0	61.13	-163.89	Breeding	2005 – 2006	
Canvasback Lake	anvasback Lake Western 0 0 0		0	0	7	7	66.39	-146.36	Breeding	1999 – 2002		
Orca Inlet, Gulf of Alaska	Western	0	0	0	0	20	12	60.55	-145.80	Spring migration	2000	
Juneau	Western	0	0	34	12	19	10	58.21	-134.27	Winter	2001 2008 – 2019	
Kodiak Island	Western	22	15	0	0	0	0	57.73	-152.50	Winter	2004 – 2006	
Montague Strait, Prince William Sound	Western	0	0	12	5	0	0	60.36	-147.22	Spring migration	1998	
Yakutat Bay	Western	0	0	8	4	7	3	59.63	-139.72	Spring migration	2019	

Fox Islands, Aleutian Islands	Western	10	6	0	0	0	0	53.85	-166.57	Winter	2007
Nelson Lagoon, Aleutian Islands	Western	34	25	0	0	0	0	56.00	-161.08	Spring migration	2003 – 2004 2015 – 2016
BRITISH COLUMBIA											
Queen Charlotte Sound	Western	0	0	28	18	0	0	53.93	-128.72	Spring migration	2014
Vancouver Island	Western	15	10	9	6	26	11	49.30	-124.50	Winter	2003 – 2005
BAJA CALIFORNIA											
Baja California	Western	0	0	15	7	0	0	30.39	-115.97	Winter	2005 – 2006
CALIFORNIA											
San Francisco Bay	Western	0	0	29	15	0	0	37.72	-122.26	Winter	2003 – 2006
WASHINGTON STATE											
Puget Sound	Western	0	0	32	29	19	17	47.50	-122.40	Winter	2003 – 2006
GRAND TOTAL		213	160	374	235	195	143				

Notes: BS = black scoter (Melanitta americana), SS = surf scoter (Melanitta perspicillata), W-WS = white-winged scoter (Melanitta deglandi); Dep = deployed,

Ret = retained; Lat = latitude; Long = longitude; Dep season = deployment season; Dep years = deployment years; Spring migr = spring migration.

JS Lamb et al. – Supporting Information

Table S2. Location and size of each cluster ($n = 88$) used by North American scoters tracked from 1998 to 2019, calculated
from an unsupervised clustering approach (bold text denotes the ten highest-ranked multispecies sites)

Cluster	Latitude	Longitude	Site name	General area	Country	Habitats	Average rank, all
ID							centrality metrics
1	55.94	-160.77	Nelson Lagoon, Aleutian Islands	SW Alaska	US	Marine	28.0
2	59.6	-162.21	Kuskokwin Bay, Bethel Census Area	SW Alaska	US	Marine	39.0
3	61.31	-165.02	Hooper Bay and Scammon Bay, Kusilvak	Alaska	US	Terrestrial	38.4
			Census Area				
4	53.75	-166.62	Unimak Bight, Aleutian Islands	SW Alaska	US	Marine	68.9
5	59.82	-163.81	Etolin Strait (Cook Strait)	SW Alaska	US	Marine	20.6
6	55.41	-162.86	Bechevin Bay, Aleutian Islands	SW Alaska	US	Marine	55.3
7	58.75	-160.31	Togiak Bay, Iilgayaq (Bristol Bay)	SW Alaska	US	Marine	63.9
8	65.29	-161.48	Kotzebue Sound and Norton Sound, NW Arctic	NW Alaska	US	Terrestrial	75.1
			Borough and Nome Census Area				
9	49.86	-124.98	Strait of Georgia	Vancouver Island, BC	Canada	Marine	11.9
10	54.06	-131.59	Hecate Strait	Haida Gwaii, BC	Canada	Marine	16.4
11	58.41	-157.64	Kvichak Bay and Nushagak Bay, Dillingham	SW Alaska	US	Marine	7.3
			Census Area				
12	53.54	-129.12	Queen Charlotte Sound	Kitimat, BC	Canada	Marine	50.9
13	49.17	-122.88	Salish Sea	Whatcom County,	US/	Marine	21.1
				Washington State/	Canada		
				Vancouver, BC			
14	59.94	-139.98	Yakutat Bay	SE Alaska	US	Marine	59.6
15	60.44	-146.44	Montague Strait, Prince William Sound	Southcentral Alaska	US	Marine	34.9
16	59.24	-154.11	Cook Inlet (Kamishak Bay, Cungaaciq), Kenai	Southcentral Alaska	US	Marine	48.5
			Peninsula				
17	56.88	-158.91	Aniakchak, Iilgayaq (Bristol Bay)	SW Alaska	US	Marine	63.6
18	57.81	-152.49	Kodiak Island and Kenai Fjords	Southcentral Alaska	US	Marine	49.5
19	47.61	-122.99	Puget Sound	Washington State	US	Marine	10.5
20	61	-107.11	Arctic Ocean Watershed	Northwest Territories/	Canada	Terrestrial	32.4
				Saskatchewan			

21	60.79	-116.53	Great Slave Lake (W side), Arctic Ocean	Northwest Territories/	Canada	Terrestrial	36.8
			Watershed	Alberta			
22	33.19	-117.61	Baja California	California/Baja	US/	Marine	83.9
				California	Mexico		
23	37.6	-122.17	San Francisco Bay	California	US	Marine	76.6
24	62.49	-119.55	Willow Lake, Arctic Ocean Watershed	Northwest Territories	Canada	Terrestrial	42.8
25	57.99	-134.1	Taku River, Juneau	Alaska/BC/Yukon	US/	Marine	12.6
					Canada		
26	58.47	-121.72	Peace River, Arctic Ocean Watershed	BC/Alberta/	Canada	Terrestrial	70.2
				Northwest Territories			
27	66.74	-124.74	K'áhbamítúé (Colville Lake), Arctic Ocean	Northwest Territories	Canada	Terrestrial,	21.7
			Watershed			marine	
28	68.09	-132.63	Inuvik, Mackenzie River, Arctic Ocean	Northwest Territories/	Canada	Terrestrial,	47.1
			Watershed	Yukon (Gwich'in		marine	
				Lands)			
29	68.03	-128.36	Fort Good Hope (Charter Community of K'asho	Northwest Territories	Canada	Terrestrial,	34.3
			Got'ine), Arctic Ocean Watershed			marine	
30	56.42	-132.11	Zimovia Strait and Stikine River	SE Alaska/BC	US/	Marine	53.9
					Canada		
31	54.8	-113.61	Lesser Slave Lake, Arctic Ocean Watershed	Alberta/Saskatchewan	Canada	Terrestrial	57.6
32	59.45	-136.08	Glacier Bay and Kusawa Lake	Alaska/BC/Yukon	US/	Marine	55.4
					Canada		
33	58.95	-97.69	Tadoule Lake, Hudson Bay Watershed	Manitoba/Nunavut	Canada	Terrestrial	42.2
34	59.02	-94.86	W Hudson Bay	Manitoba/Nunavut	Canada	Terrestrial	32.7
35	64.67	-116.53	Ingray Lake, Arctic Ocean Watershed	Northwest Territories	Canada	Terrestrial	74.3
36	60.4	-128.2	Watson Lake, Arctic Ocean Watershed	SE Yukon	Canada	Terrestrial	81.6
37	61.57	-111.78	Lake Claire and Great Slave Lake, Arctic Ocean	Alberta/	Canada	Terrestrial	28.8
			Watershed	Northwest Territories			
38	42.47	-124.26	Coos Bay	N California/	US	Marine	61.9
				Oregon			
39	61.51	-102.59	Dubawnt Lake, Hudson Bay Watershed	Northwest Territories/	Canada	Terrestrial	45.5
				Nunavut/Saskatchewan/			
				Manitoba			

40	66.11	-120.74	Sahtú (Great Bear Lake), Arctic Ocean Watershed	Northwest Territories	Canada	Terrestrial	52.6
41	67.16	-139.55	Mackenzie Bay, Arctic Ocean and Pacific Ocean	Northeastern	US/	Terrestrial	63.8
			Watersheds	Alaska/Yukon	Canada		
42	66.62	-146.21	Yukon River, Pacific Ocean Watershed	Alaska	US	Terrestrial	72.3
43	55.31	-86.48	Winisk River, Hudson Bay	Ontario	Canada	Terrestrial	60.6
44	56.32	-79.34	SE Hudson Bay	Sanikiluaq Inuit Land	Canada	Terrestrial	61.5
45	41.55	-70.02	Cape Cod Bay and Eastern Nantucket Sound	Massachusetts	US	Marine	6.7
46	40.29	-73.8	Lower Hudson River and Barnegat Bay	New York/New Jersey	US	Marine	63.3
47	50.05	-62.38	Northern Gulf of St Lawrence	Quebec	Canada	Marine	70.6
48	51.69	-80.64	Moose River, Hudson Bay Watershed	SW James Bay	Canada	Terrestrial	34.6
49	54.21	-79.44	Akimiski Island, Hudson Bay Watershed	Nunavut	Canada	Terrestrial	15.5
50	41.24	-70.41	W Nantucket Sound and Buzzards Bay	Massachusetts	US	Marine	3.3
51	38.96	-74.98	Delaware Bay	New Jersey/Delaware	US	Marine	20.8
52	46.41	-64.5	Northumberland Strait, Gulf of St Lawrence and Bay of Fundy	New Brunswick	Canada	Marine	37.5
53	56.98	_92 53	Nelson River, Hudson Bay	Manitoba	Canada	Terrestrial	61.1
54	56.85		SW Hudson Bay	Manitoba/Ontario	Canada	Terrestrial	39.8
55	51.89	_79.55	Hannah Bay, Hudson Bay, Watershed	Ontario/Quebec	Canada	Terrestrial	19.5
56	41.13	-71 79	Block Island Sound	Rhode Island/New	US	Marine	96
50	11.12	, 1.,)	Diotri Iolana Souna	York	00	101411110	<i></i>
57	48.11	-66.25	Chaleur Bay, Gulf of St Lawrence	New	Canada	Marine	26.0
				Brunswick/Quebec			
58	47.18	-64.85	Miramichi Bay, Gulf of St Lawrence	New Brunswick	Canada	Marine	26.8
59	59.04	-78.1	E Hudson Bay	Quebec	Canada	Terrestrial	81.6
60	56.76	-76.59	Umiujaq, Hudson Bay	Quebec	Canada	Terrestrial	44.8
61	31.83	-80.88	Blake Plateau	Georgia/South Carolina	US	Marine	71.6
62	44.25	-68.58	Penobscot Bay	Maine	US	Marine	61.4
63	48.4	-69.2	Upper St Lawrence River	Quebec	Canada	Marine	2.8
64	52.37	-78.82	SE James Bay, Hudson Bay Watershed	Quebec	Canada	Terrestrial	30.3
65	44.73	-66.49	Grand Manan Basin, Gulf of Maine	Maine/New Brunswick	US/	Marine	41.8
					Canada		
66	42.88	-70.71	Massachusetts Bay and Casco Bay	Massachusetts/New	US	Marine	45.0
				Hampshire/Maine			

67	50.11	-65.73	W Gulf of St Lawrence	Quebec	Canada	Marine	14.7
68	52.85	-80.87	James Bay, Hudson Bay Watershed	Ontario	Canada	Terrestrial	26.2
69	40.84	-72.72	Long Island Sound	New York/Connecticut	US	Marine	30.5
70	37.94	-76.03	Lower Chesapeake Bay	Maryland/Virginia	US	Marine	29.0
71	49.18	-67.85	Upper St Lawrence Estuary	Quebec	Canada	Marine	18.6
72	54.14	-82.35	NW James Bay, Hudson Bay Watershed	Ontario	Canada	Terrestrial	45.7
73	37.21	-75.93	Mouth of Chesapeake Bay	Virginia	US	Marine	43.7
74	53.86	-58.42	Hamilton Bank, Labrador Sea	Newfoundland/Labrado	Canada	Marine	67.6
				r			
75	56.14	-61.36	Saglek Bank, Labrador Sea	Newfoundland/Labrado	Canada	Marine	55.5
				r			
76	56.33	-68.41	Ungava Bay	Quebec	Canada	Terrestrial,	65.2
						marine	
77	56.67	-72.51	Clearwater Lakes (Lac à l'Eau Claire)	Quebec	Canada	Terrestrial	77.0
78	35.28	-76.33	Pamlico Sound	North Carolina	US	Marine	50.2
79	38.57	-76.26	Upper Chesapeake Bay	Washington, DC, and	US	Marine	54.9
				Maryland			
80	48.39	-64.65	Honguedo Strait, Gulf of St Lawrence	Quebec	Canada	Marine	21.6
81	48.88	-68.6	Lower St Lawrence Estuary	Quebec	Canada	Marine	12.9
82	33.05	-79.26	Long Bay	North Carolina/South	US	Marine	70.1
				Carolina			
83	44.04	-76.85	Lake Ontario	Ontario	Canada	Terrestrial	54.8
84	46.15	-63.58	Northumberland Strait, Gulf of St Lawrence	Nova Scotia/	Canada	Marine	24.9
				Prince Edward Island			
85	46.55	-60.76	Cabot Strait, Gulf of St Lawrence	Nova Scotia	Canada	Marine	79.2
86	52.77	-72	Lake Mistassini (Lac Mistassini), Hudson Bay	Quebec	Canada	Terrestrial	53.1
			Watershed and Atlantic Ocean Watershed				
87	53.97	-64.64	George River	Quebec/	Canada	Terrestrial	77.9
				Newfoundland &			
				Labrador			
88	43.16	-80.06	Lake Erie	Ohio	US	Terrestrial	61.8